ORIGINAL ARTICLE

Insulin resistance and adverse lipid profile in untreated very early rheumatoid arthritis patients: A single-center, cross-sectional study in China

Lu Ye¹^(b), Xin Zhang¹^(b), Huaxiang Wu¹^(b), Yahui Chen¹^(b), Haibo Zhou²^(b), Qiaohong Wang¹^(b), Weihong Xu¹^(b)

¹Department of Rheumatology, School of Medicine, The Second Affiliated Hospital of Zhejiang University, Hangzhou, China ²Department of Gastroenterology, School of Medicine, The Second Affiliated Hospital of Zhejiang University, Hangzhou, China

ABSTRACT

Objectives: This study aims to evaluate the presence and factors related to insulin resistance (IR) in untreated very early rheumatoid arthritis (RA) patients.

Patients and methods: Between June 2020 and July 2021, a total of 90 RA patients (29 males, 61 females; mean age: 49.3 ± 10.2 years; range 24 to 68 years) and 90 age-, sex- and body mass index (BMI)-matched controls (35 males, 55 females; mean age: 48.3 ± 5.1 years; range 38 to 62 years) were included. Homeostatic model assessment was applied to evaluate IR (HOMA-IR) and β -cell function (HOMA- β). Disease activity score 28 (DAS28) was used to estimate disease activity. Lipid profile, hemoglobin A1c (HbA1c), glucose, insulin, C-reactive protein (CRP), and erythrocyte sedimentation rate (ESR) were measured. Logistic regression analysis was performed to investigate the relationship between the IR and clinical features of RA patients.

Results: The RA patients had higher HOMA-IR values (p<0.001) and adverse lipid profile. The IR was positively correlated with age (r=0.35, p<0.01), CRP (r=0.42, p<0.001), ESR (r=0.33, p<0.01), disease duration (r=0.28, p<0.01), and DAS28 (r=0.50, p<0.001). The DAS28, CRP and age, but not sex and menopausal status, were independently associated with IR.

Conclusion: Insulin resistance was present in untreated very early RA patients. The DAS28, CRP, and age were independent predictors for the presence of IR. Based on these findings, RA patients should be evaluated early for the presence of IR to reduce the risk of metabolic diseases. *Keywords:* Abnormal lipid level, disease activity, insulin resistance, very early rheumatoid arthritis.

Rheumatoid arthritis (RA) is a chronic autoimmune disease with the presence of multiple autoantibodies and systemic inflammation that may lead to progressive synovitis, bone erosion and destruction of the joints, which is associated with an increased risk of morbidity and mortality. The prevalence of metabolic syndrome characterized by obesity, increased blood pressure, dyslipidemia and hyperglycemia has been enormously increasing worldwide, in part, linked to the epidemic of RA.¹⁻³ Insulin resistance (IR) status is a core feature of metabolic syndrome, and if left uncontrolled, may lead to type 2 diabetes and cardiovascular disease. Chronic low-grade inflammation involved in the pathogenesis of IR has been delineated.⁴⁻⁶ Inflammatory cytokines including tumor necrosis factor-alpha (TNF- α) and interleukin (IL)-6, various adipokines and acute-phase reactants are involved in the development of IR, which indicates inflammatory disease such as RA may be a potential cause for metabolic diseases. This

Correspondence: Weihong Xu, MD. Department of Gastroenterology, School of Medicine, The Second Affiliated Hospital of Zhejiang University, 310000 Hangzhou, China. Tel: 86-0571-8971-3754 e-mail: drwennyxu@zju.edu.cn

Citation:

Ye L, Zhang X, Wu H, Chen Y, Zhou H, Wang Q, et al. Insulin resistance and adverse lipid profile in untreated very early rheumatoid arthritis patients: A singlecenter, cross-sectional study in China. Arch Rheumatol 2022;37(4):593-602.

©2022 Turkish League Against Rheumatism. All rights reserved.

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes (http://creativecommons.org/licenses/by-nc/4.0/).

Received: August 13, 2021 Accepted: January 03, 2022 Published online: June 06, 2022

is further supported by a recent study showing a rapid beneficial effect on IR and insulin sensitivity in non-diabetic RA patients via IL-6 receptor blockade.⁷

Several studies have reported the presence of IR and impaired β -cell function in RA patients. However, most of these studies involved subjects with long-term treated RA.8-10 The potential effects of RA medication on IR may be overlooked in these studies. Indeed, although the effect of non-steroidal anti-inflammatory drugs (NSAIDs) on IR is guite limited,⁸ the use of glucocorticoids obviously alters glucose metabolism. Dessein and Joffe⁹ reported that short-term treatment with low daily dose glucocorticoids resulted enhanced β -cell function due to its in anti-inflammatory effects, while another study indicated that chronic glucocorticoid use might lead to decreased insulin sensitivity, impaired glucose tolerance (IGT), and β -cell function in RA patients.¹¹ Moreover, disease-modifying anti-rheumatic drugs (DMARDs) and anti-TNF- α therapy significantly reduced homeostasis model assessment of IR (HOMA-IR)¹² and the incidence of metabolic syndrome.¹³

Little is known about whether IR occurs at the onset of the disease or due to chronic longstanding systemic inflammation or drug exposure such as glucocorticoids.9,10,14 Moreover, data on sex-specific variations and menopausal status and their impact on IR in early RA patients are rare. Sex-related variations and menopausal status have not been described in treatment-naive very early RA patients. To address this, in the present study, we aimed to investigate the presence and factors associated with IR in untreated very early RA subjects.

PATIENTS AND METHODS

This single-center, cross-sectional study was conducted at School of Medicine, the Second Affiliated Hospital of Zhejiang University and health checkup center, Department of Rheumatology between June 2020 and July 2021. A total of 90 RA patients (29 males, 61 females; mean age: 49.3±10.2 years; range 24 to 68 years) and 90 age-, sex- and body mass index (BMI)-matched controls (35 males, 55 females; mean age: 48.3 ± 5.1 years; range 38 to 62 years) were recruited. The patients were evaluated when very early RA diagnosis was made. Inclusion criteria were as follows: RA disease duration was required to be <12 weeks, age ≥ 18 years, and a diagnosis of RA according to the 2010 American College of Rheumatology (ACR)/European League Against Rheumatism (EULAR) classification criteria.¹⁵ Those with chronic disease such as IGT status, diabetes, myocardial infarction, stroke, renal failure, cancer and other inflammatory disease were not included. Patients and controls were excluded. if they were pregnant or breastfeeding or had a presence of infection. To minimize the potential effects of drug exposure on IR, those receiving anti-TNF- α therapy, prednisone or DMARDs, or anti-diabetic drugs were also excluded. However, as NSAIDs are often used in the management from the initial phase of RA, patients undergoing NSAIDs were not excluded.

Diagnostic criteria

The HOMA-IR cut-off values varied in different population due to genetic, physiological and environmental factors. In the present study, a HOMA-IR score of >2.41 was defined as IR based on the 75th percentile of healthy individuals in a large Chinese cohort.¹⁶

Impaired glucose tolerance and diabetes were diagnosed according to the recommendations of the American Diabetes Association (ADA).¹⁷ All patients and controls underwent an oral glucose tolerance test (OGTT). Those with IGT status and diabetes were excluded. Normal BMI category ranged from 18.5 to 24.9 kg/m². Obesity was defined as a BMI of \geq 30 kg/m².¹⁸ Menopausal status was determined by self-report.

Assessment

The HOMA-IR developed by Matthews et al.¹⁹ was applied as a parameter to estimate IR (HOMA-IR) and β -cell function (HOMA- β).²⁰ The calculation formula was as follows: HOMA-IR=fasting insulin (μ U/ml)×FG (mmol/L) /22.5. As C-peptide is widely considered as a better marker than insulin for β -cell function,²¹ in this study, we used C-peptide to evaluate β -cell function by using the HOMA calculator. Disease activity score 28 (DAS28) using erythrocyte

sedimentation rate (ESR) was used to estimate disease activity.

General characteristics and biochemical measurements

All examinations were performed according to standard protocols. Height and weight were measured in the morning with the subjects wearing light clothing and no shoes. The BMI was calculated by dividing the individual's weight in kilograms by the square of height in meters.

Blood samples were collected after a 12-h overnight fast. The serum levels of total cholesterol (TC), triglyceride (TG) and glucose were detected by enzymatic methods with an automatic biochemical analyzer (AU-5831, Beckman Coulter, Brea, USA). Direct assay methods were used for measuring high-density lipoprotein (HDL) cholesterol and low-density lipoprotein (LDL) cholesterol by an automatic biochemical analyzer (AU-5831, Beckman Coulter, Brea, USA). Apolipoprotein A1 (APOA1) and apolipoprotein B (APOB) were also determined by an automatic biochemical analyzer (AU-5831, Beckman Coulter, Brea, USA). Hemoglobin A1C (HbA1c) was determined by an automatic glycohemoglobin analyzer (HLC-723G8, Tosoh Corp. Kyoto, Japan). Insulin and C-peptide were detected by an ADVIA Centaur XP automatic chemiluminescence system (Siemens Healthineers AG, Erlangen, Bayern, Germany). C-reactive protein (CRP) was detected by immunoturbidimetric assay with an automatic biochemical analyzer (AU-5831, Beckman Coulter, Brea, USA). The ESR was determined with an automatic method (Alifax Test 1, Alifax, Padova, Italy). Rheumatoid factor (RF) was detected by an automatic specific protein analyzer (Siemens Healthcare, Erlangen, Bayern, Germany). Anti-cyclic citrullinated peptide (anti-CCP) was measured by an automatic enzyme immunoassay analyzer (Quanta lyser-240, Inova Diagnostics, San Diego, CA, USA).

Disease duration was defined from RA-associated symptom onset to diagnosis. Swollen joint count (28 joints) and tender joint count (28 joints) were examined by an experienced rheumatologist. Self-assessment of disease activity by patients was measured by a Visual Analog Scale (VAS) ranging from 0 to 100 mm. Then, the DAS28 was calculated by using the DAS28 calculator. $^{\rm 22}$

Statistical analysis

Statistical analysis was performed using the SPSS version 16.0 software (SPSS Inc., Chicago, IL, USA). Descriptive data were expressed in mean \pm standard error (SE) for continuous variables and in number and frequency for categorical variables. Values of HOMA-IR were logarithm transformed before analysis, as they were non-normally distributed. The chi-square test or one-way analysis of variance (ANOVA) was used to compare characteristics of the patient groups. For comparisons between two groups, the two-tailed Student t-test was used for normally distributed variables, and logarithm transformed before analysis or Wilcoxon rank-sum test for non-normally distributed variables. Correlation analysis was assessed by the Pearson correlation analysis. Univariate and multivariate logistic regression were performed to ascertain potential independent factors associated with the presence of IR. A p value of < 0.05 was considered statistically significant with 95% confidence interval (CI).

RESULTS

Clinical characteristics of the patients and healthy subjects

A total of 90 RA patients and 90 controls were enrolled in this study. The clinical characteristics of the subjects are shown in Table 1. Compared to the controls, RA patients had increased levels of HOMA-IR, CRP, ESR, TG, LDL cholesterol, TG/HDL cholesterol ratio and fasting insulin (p<0.001, p<0.001, p<0.001, p<0.01, p<0.05, p<0.001, and p<0.001, respectively), and decreased level of HDL and APOA1 (p<0.001 for both). There were no significant differences in the HOMA- β value, TC, APOB, glucose and C-peptide levels between the RA group and controls.

HOMA-IR value was increased in RA patients

In our study, 36% of RA subjects and 19% of the controls showed abnormal HOMA-IR

		RA patients (n=90)					Controls (n=90)				
Characteristics	n	%	Mean±SD	Range	n	%	Mean±SD	Range			
Age (year)			49.3±10.2	24-68			48.3±5.1	38-62			
Sex											
Female	61	68			55	61					
Male	29	32			35	39					
BMI (kg/m²)			22.0±2.5				22.4±1.9				
Lipid profile											
TG (mmol/L)			$1.8 \pm 0.9^{**}$				1.4±0.6				
TC (mmol/L)			4.4±1.1				4.4±1.1				
HDL (mmol/L)			1.1±0.3***				1.4 ± 0.3				
LDL (mmol/L)			$2.8 \pm 0.8^{*}$				2.6±0.6				
Apo A1 (g/L)			$1.0 \pm 0.2^{***}$				1.2 ± 0.2				
Apo B (g/L)			1.0 ± 0.2				1.0 ± 0.2				
TG/HDL ratio			$1.9 \pm 1.5^{***}$				1.2 ± 0.8				
Fasting serum glucose (mmol/L)			5.0 ± 0.5				4.9±0.7				
Fasting serum insulin (mLU/mL)			10.6±4.8***				7.2±3.8				
C-Peptide (nmol/L)			0.4 ± 0.1				0.5 ± 0.1				
HOMA-IR			2.3±1.1***				1.6 ± 1.0				
HOMA-IR >2.41	32	36			17	19					
HOMA-beta cell (%)			92.8±28.3				103.3±36.3				
HbA1c (%)			5.5 ± 0.4				5.5±0.4				
CRP (mg/L)			40.2±32.0***				2.3±2.5				
ESR (mm/h)			52.2±31.4***				8.8±5.6				

RA: Rheumatoid arthritis; SD: Standard deviation; BMI: Body mass index; TG: Triglyceride; TC: Total cholesterol; HDL: High-density lipoprotein; LDL: Low-density lipoprotein; APO A1: Apolipoprotein A1; APO B: Apolipoprotein B; HOMA: Homeostasis model assessment; IR: Insulin resistance; HbA1c: Hemoglobin A1c; CRP: C-reactive protein; ESR: Erythrocyte sedimentation rate; Comparisons between RA patients with controls are assessed by one-way ANOVA or Chi-square test. Only statistically significant differences are marked with symbols. A p value <0.05 is considered as statistically significant. * p<0.05, ** p<0.01,*** p<0.01.

values. Compared to the control group, the mean HOMA-IR value was significantly higher in RA patients (2.3 ± 1.1) than controls $(1.6\pm1.0, p<0.001)$.

Comparisons of characteristics between RA groups

The RA patients were further divided into three subgroups according to sex and menopausal status. The clinical characteristics of the subjects are shown in Table 2 and Table 3.

Compared to the premenopausal patients, male RA patients had higher TG $(2.0\pm0.9 \text{ vs.})$

1.5±0.8 mmol/L, p<0.05), LDL cholesterol (3.1±0.6 vs. 2.4±0.7 mmol/L, p<0.001), APOB (1.1±0.1 vs. 0.9±0.2 g/L, p<0.001), fasting glucose (5.0±0.5 vs. 4.8±0.4 mmol/L, p<0.05) and HOMA-IR value (2.4±1.1 vs. 2.2±1.1, p>0.05), and with longer disease duration, but lower HOMA- β value (86.8±24.8% vs. 104.1±28.8%, p<0.05) and HDL cholesterol (1.0±0.2 vs. 1.1±0.3 mmol/L, p<0.05). Compared to the premenopausal patients, postmenopausal patients had higher APOB (1.0±0.2 vs. 0.9±0.2 g/L, p<0.05), LDL cholesterol (2.9±0.8 vs. 2.4±0.7 mmol/L, p<0.05) and fasting glucose (5.0±0.5 vs. 4.8±0.4 mmol/L,

(n=29) $(n=29)$ $(n=29)$ Characteristics n % Mean±SD Range n Age (year) 50.0±10.1 24-68*** 2 24-68*** 1 BMI (kg/m ²) 21.7±2.4 21.7±2.4 2	RA ange n % 68***	atients (n=90) Mean±SD Re 38.3±5.7 27-4 21.5±2.4 21.5±0.8 4.0±1.2 1.1±0.3 2.4±0.7# 1.1±0.3	ange n % 16###	Controls (n=90) Mean±SD 55.5±6.3 22.5±2.6	Range
Characteristics n %e Mean±SD Range n Age (year) 50.0±10.1 24-68*** 50.0±10.1 24-68*** BMI (kg/m ²) 21.7±2.4 21.7±2.4 21.7±2.4 $= 1.754$ BMI (kg/m ²) 21.7±2.4 21.7±2.4 $= 1.754$ $= 21.754$ Lipid profile 21.7±2.4 21.7±2.4 $= 1.056$ $= 1.056$ TG (mmol/L) 2.0±0.9* 2.0±0.9* $= 1.056$ $= 1.056$ HDL (mmol/L) 2.0±0.2* 1.0±0.2* $= 1.066^{***}$ $= 1.066^{***}$ Apo A1 (g/L) 3.1±0.6*** $= 1.066^{***}$ $= 1.066^{***}$ $= 1.066^{***}$ Apo A1 (g/L) 1.0±0.2* $= 1.066^{***}$ $= 1.066^{***}$ $= 1.066^{***}$ Apo B (g/L) 7.0±0.2* $= 1.016^{***}$ $= 2.2±1.3$ $= 2.2±1.3$ Fasting serum isculin (mU/mL) $= 2.2±1.3$ $= 2.2±1.3$ $= 2.2±1.3$ $= 2.2±1.3$ Fasting serum insulin (mU/mL) $= 0.4\pm0.1$ $= 0.4\pm0.1$ $= 0.4\pm0.1$ $= 0.4\pm0.1$ Cybertide (mmo/L) $= 0.4\pm0$	ange n %	Mean±SD Ra 38.3±5.7 27-4 21.5±2.4 1.5±0.8 4.0±1.2 1.1±0.3 2.4±0.7# 1.1±0.3 0.0±0.2#	ange n %	Mean±SD 55.5±6.3 22.5±2.6	Range
Age (year)50.0±10.124-68***BMI (kg/m²) 21.7 ± 2.4 21.7 ± 2.4 Lipid profile $2.1.7\pm2.4$ $2.0\pm0.9^{*}$ TG (mmol/L) $2.0\pm0.9^{*}$ 4.6 ± 0.9 TC (mmol/L) $1.0\pm0.2^{*}$ $1.0\pm0.2^{*}$ HDL (mmol/L) $1.0\pm0.2^{*}$ $1.0\pm0.2^{*}$ HDL (mmol/L) $1.0\pm0.2^{*}$ $1.0\pm0.2^{*}$ Apo A1 (g/L) $1.0\pm0.2^{*}$ $1.0\pm0.2^{*}$ Apo B (g/L) $1.0\pm0.2^{*}$ $1.1\pm0.1^{**}$ TG/HDL ratio 2.2 ± 1.3 2.2 ± 1.3 Fasting serum insulin (mU/mL) 0.4 ± 0.1 0.4 ± 0.1 C-Peptide (mmol/L) 0.4 ± 0.1 0.4 ± 0.1	68**	38.3±5.7 27-4 21.5±2.4 1.5±0.8 4.0±1.2 1.1±0.3 2.4±0.7# 1.1±0.3	f6###	55.5±6.3 22.5±2.6	3
BMI (kg/m^2) 21.7 ± 2.4 Lipid profile $2.0\pm0.9^*$ TG ($mmol/L$) $2.0\pm0.9^*$ TC ($mmol/L$) 4.6 ± 0.9 HDL ($mmol/L$) $2.0\pm0.2^*$ HDL ($mmol/L$) $1.0\pm0.2^*$ HDL ($mmol/L$) $1.0\pm0.2^*$ Apo A1 (g/L) 1.0 ± 0.2 Apo A1 (g/L) 1.0 ± 0.2 Apo B (g/L) $1.1\pm0.1^{***}$ TG/HDL ratio 2.2 ± 1.3 Fasting serum glucose ($mmol/L$) $5.0\pm0.5^*$ Fasting serum insulin (mU/mL) 0.4 ± 0.1		21.5 ± 2.4 1.5 ± 0.8 4.0 ± 1.2 1.1 ± 0.3 $2.4\pm0.7\#$ 1.1 ± 0.3 $0.9\pm0.2\#$		22.5±2.6	47-67
Lipid profile $2.0\pm0.9^{\circ}$ TG (mmol/L) $2.0\pm0.9^{\circ}$ TC (mmol/L) 4.6 ± 0.9 HDL (mmol/L) $1.0\pm0.2^{\circ}$ HDL (mmol/L) $1.0\pm0.2^{\circ}$ Apo A1 (g/L) $1.0\pm0.2^{\circ}$ Apo B (g/L) $1.1\pm0.1^{***}$ TG/HDL ratio 2.2 ± 1.3 Fasting serum isulin (mIU/mL) $5.0\pm0.5^{\circ}$ Fasting serum insulin (mIU/mL) 0.4 ± 0.1		1.5 ± 0.8 4.0 ± 1.2 1.1 ± 0.3 $2.4\pm0.7\#$ 1.1 ± 0.3 $0.9\pm0.2\#$			
TG (mmol/L) $2.0\pm0.9^{*}$ TC (mmol/L) 4.6 ± 0.9 HDL (mmol/L) $1.0\pm0.2^{*}$ LDL (mmol/L) $3.1\pm0.6^{***}$ Apo A1 (g/L) 1.0 ± 0.2 Apo B (g/L) 1.0 ± 0.2 TG/HDL ratio 2.2 ± 1.3 Fasting serum glucose (mmol/L) $5.0\pm0.5^{*}$ Fasting serum insulin (mU/mL) 11.2 ± 4.9 C-Peptide (mmol/L) 0.4 ± 0.1		1.5±0.8 4.0±1.2 1.1±0.3 2.4±0.7# 1.1±0.3			
TC (mmo/L) 4.6 ± 0.9 HDL (mmo/L) $1.0\pm0.2^{*}$ LDL (mmo/L) $3.1\pm0.6^{***}$ Apo Al (g/L) 1.0 ± 0.2 Apo B (g/L) $1.1\pm0.1^{***}$ TG/HDL ratio 2.2 ± 1.3 Fasting serum glucose (mmo/L) $5.0\pm0.5^{*}$ C-Peptide (nmo/L) 0.4 ± 0.1		4.0±1.2 1.1±0.3 2.4±0.7# 1.1±0.3 0.9±0.2#		1.8 ± 1.0	
HDL (mmol/L) 1.0±0.2* LDL (mmol/L) 3.1±0.6*** Apo A1 (g/L) 1.0±0.2 Apo B (g/L) 1.1±0.1*** TG/HDL ratio 2.2±1.3 Fasting serum glucose (mmol/L) 5.0±0.5* Fasting serum insulin (mIU/mL) 0.4±0.1		1.1±0.3 2.4±0.7# 1.1±0.3 0.9+0.2#		4.6 ± 1.2	
LDL (mmol/L) 3.1±0.6*** Apo A1 (g/L) 1.0±0.2 Apo B (g/L) 1.1±0.1*** TG/HDL ratio 2.2±1.3 Fasting serum glucose (mmol/L) 5.0±0.5* Fasting serum insulin (mlU/mL) 0.4±0.1		2.4±0.7# 1.1±0.3 0 9+0 2#		1.1 ± 0.3	
Apo A1 (g/L) 1.0±0.2 Apo B (g/L) 1.1±0.1*** TG/HDL ratio 2.2±1.3 Fasting serum glucose (mmol/L) 5.0±0.5* Fasting serum insulin (mIU/mL) 11.2±4.9 C-Peptide (nmol/L) 0.4±0.1		1.1±0.3 0 9+0 2#		2.9 ± 0.8	
Apo B (g/L) 1.1±0.1*** TG/HDL ratio 2.2±1.3 Fasting serum glucose (mmol/L) 5.0±0.5* Fasting serum insulin (mlU/mL) 11.2±4.9 C-Peptide (mmol/L) 0.4±0.1		#6 U+0 U		1.1 ± 0.2	
TG/HDL ratio2.2±1.3Fasting serum glucose (mmol/L)5.0±0.5*Fasting serum insulin (mIU/mL)11.2±4.9C-Peptide (nmol/L)0.4±0.1		·····		1.0 ± 0.2	
Fasting serum glucose (mmol/L)5.0±0.5*Fasting serum insulin (mIU/mL)11.2±4.9C-Peptide (nmol/L)0.4±0.1		1.5 ± 1.2		2.0 ± 1.7	
Fasting serum insulin (mIU/mL) 11.2±4:9 C-Peptide (nmol/L) 0.4±0.1		4.8±0.4#		5.0±0.5	
C-Peptide (nmol/L) 0.4±0.1		10.3 ± 4.7		10.4 ± 4.9	
		0.4 ± 0.1		0.4 ± 0.2	
HOMA-IR 2.4±1.1		2.2±1.1		2.3 ± 1.2	
HOMA-IR >2.41 11 37.93 77 3	7 30.43		14 36.84		
HOMA-beta cell (%) 86.8±24.8*		104.2 ± 28.8		90.4 ± 28.5	
HbA1c 5.5±0.4		5.3 ± 0.5		5.5 ± 0.4	
CRP (mg/L) 46.4±34.1		44.6 ±37.3		32.1 ± 24.1	
ESR (mm/h) 49.0±32.3		54.3±32.2		53.4±30.9	

Ma	le					Table 3. Disease-specific parameters of RA subjects									
	Male		Female			Female									
(n=29)			Premenopausal group (n=23)			Postmenopausal group (n=38)									
%	Mean±SD	n	%	Mean±SD	n	%	Mean±SD								
	411.1±631.8			360.7±649.7			159.2±198.8▲								
	416.4±423.9			368.2±308.3			437.3±467.3								
	6.9±2.7			$5.5 \pm 2.0^{*}$			6.7±2.5								
	4.3±0.9			4.1±0.8			4.2±0.7								
6.90		2	8.70		3	7.89									
72.41		19	82.61		30	78.95									
20.69		2	8.70		5	13.16									
79.31		16	69.57		29	76.32									
	(n=2 % 6.90 72.41 20.69 79.31	(n=29) % Mean±SD 411.1±631.8 416.4±423.9 6.9±2.7 4.3±0.9 6.90 72.41 20.69 79.31	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	(n=29) Premenopausa % Mean±SD n % 411.1±631.8 416.4±423.9 5.9±2.7 5.9±2.7 4.3±0.9 2 8.70 72.41 19 82.61 20.69 2 8.70 79.31 16 69.57	$\begin{array}{c c c c c c c } & & & & & & & & & & & & & & & & & & &$	$\begin{array}{ c c c c c c } & \begin{tabular}{ c c c c c } & \begin{tabular}{ c c c c c } & \begin{tabular}{ c c c c c c } & \begin{tabular}{ c c c c c c c } & \begin{tabular}{ c c c c c c c } & \begin{tabular}{ c c c c c c c } & \begin{tabular}{ c c c c c c c } & \begin{tabular}{ c c c c c c c c } & \begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	$\begin{array}{ c c c c c c c c c c c c c c c c c c c$								

RA: Rheumatoid arthritis; SD: Standard deviation; RF: Rheumatoid factor; Anti-CCP: Anti-cyclic citrullinated peptide; DAS28: 28-joint disease activity score; NSAIDs: Non-steroidal anti-inflammatory drugs; Comparisons among groups are assessed by one-way ANOVA. Only statistically significant differences are marked with symbols. A *p* value <0.05 is considered as statistically significant; * p<0.05; ** p<0.01; *** p<0.001 represents statistically significant differences between male group and premenopausal group. $\Rightarrow p<0.05$; $\Rightarrow p<0.01$; $\Rightarrow p<0.001$ represents statistically significant differences between male group and postmenopausal group.

p<0.05). Compared to the postmenopausal female patients, younger male patients had higher RF levels (411.1 \pm 631.8 *vs.* 159.2 \pm 198.8 IU/mL, p<0.05). No significant differences in HOMA-IR score, lipid profile, HbA1c, glucose, insulin, CRP, ESR, and disease activity (DAS28) (p>0.05 for all) were observed among three groups.

DAS28 was associated with the presence of IR in RA

Pearson correlation analysis demonstrated that IR was positively correlated with age (r=0.35, p<0.01), CRP (r=0.42, p<0.001),

ESR (r=0.33, p<0.01), disease duration (r=0.28, p<0.01) and DAS28 (r=0.50, p<0.001), indicating a moderate to good correlation between IR and DAS28. It showed no significant correlation between HOMA-IR score and BMI, RF, or anti-CCP antibody. The associations of clinical characteristics with IR in RA patients are shown in Figures 1 and 2.

Univariate and multivariate logistic regression analyses were performed to ascertain potential independent factors associated with the presence of IR (Table 4). In the univariate model, DAS28 (6.003, CI: 1.638-22.005),



Figure 1. Association of age and BMI with insulin resistance in RA patients was analyzed by Pearson correlation analysis. Only statistically significant differences are marked with symbols. BMI: Body mass index; lg: Logarithm; IR: Insulin resistance; r: Correlation coefficient; RA: Rheumatoid arthritis.

Insulin resistance in untreated early rheumatoid arthritis patients



Figure 2. Association of RA features with insulin resistance in RA patients was analyzed by Pearson correlation analysis. Only statistically significant differences are marked with symbols.

CRP: C-reactive protein; ESR: Erythrocyte sedimentation rate; RF: Rheumatoid factor; lg: Logarithm; IR: Insulin resistance; Anti-CCP: Anti-cyclic citrullinated peptide; DAS28: 28-joint disease activity score; r: Correlation coefficient.

Table 4. Univariate and multivariate logistic regression analyses for factors associated with the presence of IR in RA patients									
		Univariate		Multivariate					
	OR	95% CI	р	OR	95% CI	р			
Age (year)	1.1	1.013-1.193	0.023	1.096	1.011-1.187	0.026			
Sex	0.577	0.137-2.431	0.453	1.617	0.389-6.73	0.509			
Menopausal status	0.552	0.013-23.739	0.757						
Body mass index	1.286	0.98-1.688	0.07	1.258	0.969-1.633	0.085			
Disease duration (weeks)	1.07	0.765-1.497	0.693						
DAS28 score	6.003	1.638-22.005	0.007	5.75	1.754-18.844	0.004			
RF (IU/mL)	1.001	0.999-1.002	0.505						
Anti-CCP (RU/mL)	0.999	0.998-1.001	0.396	0.999	0.998-1.001	0.475			
C-reactive protein	1.047	1.018-1.076	0.001	1.047	1.018-1.076	0.001			
Erythrocyte sedimentation rate	0.983	0.954-1.013	0.265	0.981	0.953-1.011	0.213			
Using NSAIDs	1.911	0.219-16.651	0.558						

IR: Insulin resistance; RA: Rheumatoid arthritis; CI: Confidence interval; DAS28: 28-joint disease activity score; RF: Rheumatoid factor; Anti-CCP: Anti-cyclic citrullinated peptide; NSAIDs, nonsteroidal anti-inflammatory drugs; Menopausal status is for female group. A p value <0.05 is considered as statistically significant.

age (1.1, CI: 1.013-1.193) and CRP (1.047, CI: 1.018-1.076) were associated with IR. In the multivariate model, DAS28 (5.75, CI: 1.754-18.844), age (1.096, CI: 1.011-1.187), and CRP (1.047, CI: 1.018-1.076) remained independently associated with IR.

DISCUSSION

In the present study, we observed increased HOMA-IR value and adverse lipid profile in very early RA patients. The DAS28, CRP and age, but not sex and menopausal status, were shown to be independently associated with IR in RA patients.

The presence of IR is demonstrated in established RA, but in the early stage of RA, the results are inconclusive. Interestingly, a study indicated that a severe insulin resistant state was present in early untreated RA patients; however, BMI was significantly higher in RA group compared to the controls in this study.²³ Mirjafari et al.²⁴ also observed a higher HOMA-IR score in early rheumatoid arthritis (ERA) patients; however, this study included patients treated with DMARDs and glucocorticoids. Another study included 46 untreated early RA patients. This study showed no significant difference in HOMA-IR and HOMA- β scores between early RA patients and controls. However, both RA patients and controls were overweight.²⁵ These studies provide new insights into understanding of the link between IR and RA. However, as overweight, obesity, cardiovascular diseases, and type 2 diabetes are well-recognized to be related with IR,^{26,27} these findings have limitations that IR state might have been overestimated and may be not consistent with each other and the present study.

In the current study, we applied the HOMA method to access IR and β -cell function. Hyperinsulinemic euglycemic clamp and hyperglycemic clamp are considered as the goldstandard method for quantifying insulin action and insulin secretion of β -cell, respectively. However, glucose clamp technique requires glucose readings every 5 min for a few hours, making it difficult to recruit patients in clinical and epidemiological studies. The HOMA method developed by Matthews et al.¹⁹ is a validated, simple and reliable tool to access IR and β -cell function, and is widely used in clinical study. Moreover, the close correlation between the results from HOMA and glucose clamp method is demonstrated,²⁰ making HOMA model a suitable tool for clinical study. In our study, although the mean score of HOMA-IR in RA cases was normal (mean HOMA-IR=2.33), it was statically significantly higher than controls. More intriguingly, 36% of the RA patients showed IR with a HOMA-IR score of >2.41 compared to 19% in controls in our study. A possible explanation for elevated HOMA-IR value in early RA is that various proinflammatory cytokines are demonstrated to be increased during a prolonged period of asymptomatic stage. Indeed, Karlson et al.²⁸ reported that IL-6 level was significantly higher in preclinical RA, and TNF receptor 2, a crucial inflammatory biomarker was increased 12 years prior to RA symptoms. Both IL-6 and TNF- α , core proinflammatory cytokines in clinical phase of RA, are well-known to induce IR state. Therefore, IR state may start in early RA, or even in the preclinical phase.

In addition, disruption of lipid profiles characterized by high TG and LDL cholesterol, as well as low HDL cholesterol and APOA1 were observed in RA cases in current study. The TG/HDL ratio, which is an independent risk for cardiovascular disease, was demonstrated to be elevated in the present study. These data are similar to the previous studies in early untreated RA subjects. Importantly, high TG and LDL cholesterol, along with low HDL cholesterol and APOA1, have been demonstrated as risk factors for cardiovascular disease.29,30 Recently, a study emphasized the importance of diabetes mellitus and cardiovascular risk management in patients with RA, and moreover, the authors pointed out that experience could be learnt from cardiovascular disease prevention programs to benefit RA patients.³¹

Similar to the findings in previous reports, DAS28 and CRP were found to be positively correlated with IR,32 and demonstrated to be independent predictors in logistic regression analysis in the present study. A positive association between ESR and IR was observed, but it was not an independent risk factor for IR, maybe due to relatively small sample size. However, as DAS28 is more accurate than CRP and ESR as a disease activity measure in RA, the present study confirmed disease activity as an independent predictor for the risk of IR. These findings emphasize the importance of evaluation of metabolic disorders in RA patients and early treatment of RA to reduce disease activity and prevent the development of IR and dyslipidemia.

Additionally, we also found that age was an independent risk factor for IR in RA. As the IR prevalence increases in the general population with age, it is not surprising that older male patients and postmenopausal women had higher HOMA-IR value, LDL cholesterol, APOB, and fasting glucose than premenopausal women in our study. Menopausal state is known to cause weight gain, IR, and dyslipidemia in women.³³

However, in the present study, it was not an independent risk factor for IR in RA. The possible reason may be that age of menopause is affected by patient's general health status, as reduced ovarian function has been observed in patients with chronic disease including type 2 diabetes and RA.^{34,35} Therefore, RA per se or disease activity may be a major cause for IR in these patients.

Moreover, the increased fasting insulin was observed in RA group, with no significant difference in HOMA- β value compared with controls. This result is not consistent with previous reports in established RA.^{10,11} One possible explanation for this finding is pancreatic β -cell compensation. Indeed, the changes in β -cell function in the progression of diabetes can be viewed as five stages: compensation, stable adaption, early unstable decompensation, stable decompensation and severe decompensation.³⁶ The authors indicated that, during the compensation stage, insulin secretion increased to maintain normoglycemia secondary to IR.

Nonetheless, certain limitations of the current study should be recognized. The sample size of our study is small and from a single center. Additionally, we did not have enough cases of RA patients with different genetic background and environmental factors.

In conclusion, IR was present in untreated very early RA patients in the current study. The DAS28, CRP, and age were significant independent predictors for the presence of IR. Therefore, the IR status should be evaluated early in RA patients to reduce the risk of metabolic diseases.

Ethics Committee Approval: The study protocol was approved by the School of Medicine, The Second Affiliated Hospital of Zhejiang University (IR2020001179). The study was conducted in accordance with the principles of the Declaration of Helsinki.

Patient Consent for Publication: A written informed consent was obtained from each patient.

Data Sharing Statement: The data that support the findings of this study are available from the corresponding author upon reasonable request.

Author Contributions: Statistical analysis and manuscript writing: Y.L.; study design, interpretation of data: X.W.H.; statistical analysis and interpretation of data; W.H.X.; data collection and statistical analysis: Z.X., C.Y.H.; data collection, statistical analysis and interpretation of data: Z.H.B., W.Q.H.; All authors approved the final manuscript as submitted and agree to be accountable for all aspects of the work.

Conflict of Interest: The authors declared no conflicts of interest with respect to the authorship and/or publication of this article.

Funding: This work was supported by Zhejiang Provincial Natural Science Foundation of China (grant number LQ15H070003).

REFERENCES

- Costa NT, Scavuzzi BM, Iriyoda TMV, Lozovoy MAB, Alfieri DF, de Medeiros FA, et al. Metabolic syndrome and the decreased levels of uric acid by leflunomide favor redox imbalance in patients with rheumatoid arthritis. Clin Exp Med 2018;18:363-72.
- Zaragoza-García O, Navarro-Zarza JE, Maldonado-Anicacio JY, Castro-Alarcón N, Rojas IP, Guzmán-Guzmán IP. Hypertriglyceridaemic waist is associated with hyperuricaemia and metabolic syndrome in rheumatoid arthritis patients. Diabetes Metab Syndr 2019;13:722-9.
- Mochizuki T, Ikari K, Yano K, Okazaki K. Fiveyear incidence of common comorbidities, such as hypertension, dyslipidemia, diabetes mellitus, cardiovascular disease, cerebrovascular disease and cancer, in older Japanese patients with rheumatoid arthritis. Geriatr Gerontol Int 2019;19:577-81.
- Asghar A, Sheikh N. Role of immune cells in obesity induced low grade inflammation and insulin resistance. Cell Immunol 2017;315:18-26.
- Matulewicz N, Karczewska-Kupczewska M. Insulin resistance and chronic inflammation. Postepy Hig Med Dosw (Online) 2016;70:1245-58.
- Cruz KJC, de Oliveira ARS, Morais JBS, Severo JS, Marreiro PhD DDN. Role of microRNAs on adipogenesis, chronic low-grade inflammation, and insulin resistance in obesity. Nutrition 2017;35:28-35.
- Castañeda S, Remuzgo-Martínez S, López-Mejías R, Genre F, Calvo-Alén J, Llorente I, et al. Rapid beneficial effect of the IL-6 receptor blockade on insulin resistance and insulin sensitivity in nondiabetic patients with rheumatoid arthritis. Clin Exp Rheumatol 2019;37:465-73.
- Wasko MC, Kay J, Hsia EC, Rahman MU. Diabetes mellitus and insulin resistance in patients with rheumatoid arthritis: Risk reduction in a chronic inflammatory disease. Arthritis Care Res (Hoboken) 2011;63:512-21.
- Dessein PH, Joffe BI. Insulin resistance and impaired beta cell function in rheumatoid arthritis. Arthritis Rheum 2006;54:2765-75.
- Ferraz-Amaro I, García-Dopico JA, Medina-Vega L, González-Gay MA, Díaz-González F. Impaired beta

cell function is present in nondiabetic rheumatoid arthritis patients. Arthritis Res Ther 2013;15:R17.

- Hoes JN, van der Goes MC, van Raalte DH, van der Zijl NJ, den Uyl D, Lems WF, et al. Glucose tolerance, insulin sensitivity and β-cell function in patients with rheumatoid arthritis treated with or without low-to-medium dose glucocorticoids. Ann Rheum Dis 2011;70:1887-94.
- Dessein PH, Joffe BI, Stanwix AE. Effects of disease modifying agents and dietary intervention on insulin resistance and dyslipidemia in inflammatory arthritis: A pilot study. Arthritis Res 2002;4:R12.
- 13. Toms TE, Panoulas VF, John H, Douglas KM, Kitas GD. Methotrexate therapy associates with reduced prevalence of the metabolic syndrome in rheumatoid arthritis patients over the age of 60- more than just an anti-inflammatory effect? A cross sectional study. Arthritis Res Ther 2009;11:R110.
- Chung CP, Oeser A, Solus JF, Gebretsadik T, Shintani A, Avalos I, et al. Inflammation-associated insulin resistance: Differential effects in rheumatoid arthritis and systemic lupus erythematosus define potential mechanisms. Arthritis Rheum 2008;58:2105-12.
- Aletaha D, Neogi T, Silman AJ, Funovits J, Felson DT, Bingham CO 3rd, et al. 2010 Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. Arthritis Rheum 2010;62:2569-81.
- Lu J, Bi Y, Wang T, Wang W, Mu Y, Zhao J, et al. The relationship between insulin-sensitive obesity and cardiovascular diseases in a Chinese population: Results of the REACTION study. Int J Cardiol 2014;172:388-94.
- American Diabetes Association. 2. Classification and diagnosis of diabetes: Standards of medical care in diabetes-2018. Diabetes Care 2018;41(Suppl 1):S13-S27.
- Obesity: Preventing and managing the global epidemic. Report of a WHO consultation. World Health Organ Tech Rep Ser 2000;894:i-xii, 1-253.
- Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: Insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. Diabetologia 1985;28:412-9.
- Wallace TM, Levy JC, Matthews DR. Use and abuse of HOMA modeling. Diabetes Care 2004;27:1487-95.
- Gonzalez-Mejia ME, Porchia LM, Torres-Rasgado E, Ruiz-Vivanco G, Pulido-Pérez P, Báez-Duarte BG, et al. C-peptide is a sensitive indicator for the diagnosis of metabolic syndrome in subjects from Central Mexico. Metab Syndr Relat Disord 2016;14:210-6.
- 22. Prevoo ML, van 't Hof MA, Kuper HH, van Leeuwen MA, van de Putte LB, van Riel PL. Modified disease activity scores that include twenty-eight-joint counts. Development and validation in a prospective

longitudinal study of patients with rheumatoid arthritis. Arthritis Rheum 1995;38:44-8.

- 23. Shahin D, Eltoraby E, Mesbah A, Houssen M. Insulin resistance in early untreated rheumatoid arthritis patients. Clin Biochem 2010;43:661-5.
- 24. Mirjafari H, Farragher TM, Verstappen SM, Yates A, Bunn D, Marshall T, et al. Seropositivity is associated with insulin resistance in patients with early inflammatory polyarthritis: Results from the Norfolk Arthritis Register (NOAR): An observational study. Arthritis Res Ther 2011;13:R159.
- Manrique-Arija S, Ureña I, Valdivielso P, Rioja J, Jiménez-Núñez FG, Irigoyen MV, et al. Insulin resistance and levels of adipokines in patients with untreated early rheumatoid arthritis. Clin Rheumatol 2016;35:43-53.
- Ruderman NB, Carling D, Prentki M, Cacicedo JM. AMPK, insulin resistance, and the metabolic syndrome. J Clin Invest 2013;123:2764-72.
- 27. Reaven GM. Insulin resistance: The link between obesity and cardiovascular disease. Med Clin North Am 2011;95:875-92.
- 28. Karlson EW, Chibnik LB, Tworoger SS, Lee IM, Buring JE, Shadick NA, et al. Biomarkers of inflammation and development of rheumatoid arthritis in women from two prospective cohort studies. Arthritis Rheum 2009;60:641-52.
- 29. Wang J, Shi L, Zou Y, Tang J, Cai J, Wei Y, et al. Positive association of familial longevity with the moderate-high HDL-C concentration in Bama Aging Study. Aging (Albany NY) 2018;10:3528-40.
- Chistiakov DA, Orekhov AN, Bobryshev YV. ApoA1 and ApoA1-specific self-antibodies in cardiovascular disease. Lab Invest 2016;96:708-18.
- Semb AG, Rollefstad S, Ikdahl E, Wibetoe G, Sexton J, Crowson C, et al. Diabetes mellitus and cardiovascular risk management in patients with rheumatoid arthritis: An international audit. RMD Open 2021;7:e001724.
- 32. Dao HH, Do QT, Sakamoto J. Increased frequency of metabolic syndrome among Vietnamese women with early rheumatoid arthritis: A cross-sectional study. Arthritis Res Ther 2010;12:R218.
- 33. Hallajzadeh J, Khoramdad M, Izadi N, Karamzad N, Almasi-Hashiani A, Ayubi E, et al. Metabolic syndrome and its components in premenopausal and postmenopausal women: A comprehensive systematic review and meta-analysis on observational studies. Menopause 2018;25:1155-64.
- 34. Isik S, Ozcan HN, Ozuguz U, Tutuncu YA, Berker D, Alimli AG, et al. Evaluation of ovarian reserve based on hormonal parameters, ovarian volume, and antral follicle count in women with type 2 diabetes mellitus. J Clin Endocrinol Metab 2012;97:261-9.
- 35. Brouwer J, Dolhain RJEM, Hazes JMW, Visser JA, Laven JSE. Reduced ovarian function in female rheumatoid arthritis patients trying to conceive. ACR Open Rheumatol 2019;1:327-35.
- 36. Weir GC, Bonner-Weir S. Five stages of evolving beta-cell dysfunction during progression to diabetes. Diabetes 2004;53 Suppl 3:S16-21.